# An Updated Overview on Pharmacognostical and Pharmacological Screening of *Tecoma Stans*

K. Gopalasathees Kumar\*, T. Boopathi KMCH college of Pharmacy, Coimbatore, Tamil Nadu, India gskpungai@gmail.com

#### ABSTRACT

The using of natural plants in treatment purposes are the nowadays most familiar than synthetic products because synthetic drugs can cause many of the side effects and the adverse effect. The *Tecoma stans* are one of the plants which are available in most of the tropical countries. In this plant is already used in the traditional medicine in some of the countries like India, Pakistan, etc., this plant belongs to the family of *Bignoniaceae*. This plant is having the many of the active chemical constituents and pharmacological effects. Many of the researchers are studied the pharmacological screening and the current research is going in that plant. The aim of this review was the updated research collections of this plant for its pharmacological screening. The review the on various researchers like pharmacognostical study and the *in vitro* and *in vivo* screening of various parts of *Tecoma stans*. In this review was concluded that the various parts of the plants are having various pharmacological actions like anti-inflammatory, analgesic, anticancer cardio-protective effect, genotoxic, cytotoxicity, wound healing, anti-hyperglycemic, protect CNS, gastric ulcer healing, antiproliferative, antioxidant, anti-microbial, hemolytic activity, anti-lipoxygenase and acetyl-cholinesterase inhibitory activities. And this review was used to develop the future research on this plant.

Key Words: Tecoma stans, pharmacology, pharmacognostical, anti-microbial, anti-cancer, anti-diabetic.

#### INTRODUCTION

The use of natural components for therapeutic properties is earliest as human evolution and, for a long time, plant, mineral, and animal produces the main foundations of drugs. In current years, there has been increasing attention in alternative treatments and the healing use of natural products, particularly those derived from plant life. (Divya et al., 2014, Boopathi et al., 2017, Sunita et al., 2016) This attention in drugs of plant source is due to numerous explanations, namely, conventional medicine can be unsuccessful (e.g. side effects, adverse effects and ineffective therapy), offensive or improper use of synthetic drugs result in side effects and other complications. The Indian sub-continent encompassing of the nation's India, Pakistan, and Bangladesh is the site of one of the eldest evolutions, and it has seen the growth of many traditional healthcare methods. Their growth was maintained by the great biodiversity in plants and biota due to differences in geography and weather. The Bignoniaceae family including of about 110 types and 650 species is a family of flowering plants,

normally identified as the Jacaranda family, Trumpet Creeper family, Bignonia family, or the Catalpa family. Plant species belonging to this family are dispersed universal, but most of them occur in the tropical and sub-tropical countries. However, a number of moderate species also grow in North America and East Asia. Although the family is small, the Bignoniaceae plants are significant for their stated bio-active ingredients and diverse pharmacological activities. (Anburaj et al., 2016, Raju et al., 2011) Bignoniaceae family plants are also extensively used in traditional therapeutic systems of a number of kingdoms, where traditional and tribal medicinal practitioners use a number of species for an action of diverse illnesses. This review purpose is the pharmacognostical, phytochemical profiles and therapeutic potential of different parts of Tecoma stans. (Sunitha et al., 2016)

#### Description of Tecoma stans

Tecoma stans are the belongs to the family of Bignoniaceae that is having many synonyms and common names. Synonyms of Tecoma stans are,

39

Bignonia stans, Stenolobium stans, Gelseminum stans, Kuntze Seem and Common names are Yellow Bells, Yellow Trumpet Bush, Ginger-Thomas, Yellow Elder and Esperanza. This plant is the official flower for US Virgin Islands and home flower of the Bahamas. Tecoma stan is a small tree, 5-8 m in height. The bark of the plant is pale brown to grey color; Leaves are conflicting, compound and imparipinnate with 2 to 5 pairs of leaflets and a greater single terminal leaflet.(Thirumal et al., 2013, Namde et al., 2014) Leaflets are lanceolate, up to 10 cm long, with serrated borders, mid-green above and soft to the touch. At the ends of the divisions, flowers occur in clusters and are trumpet-shaped with 5 round lobes, 6 cm lengthy, pale to bright yellow, with faint orange stripes at the throat. Fruits are narrow, slightly flattened to pointy capsules, up to 20 cm long, comprising many aerial seeds; green when young, pale brown on maturing and remain on the tree in untidy clusters for many months. (Sarfaraj et al., 2010)

# PHARMACOGNOSTICAL STUDY

In a pharmacognostical study, this plant is undergoing the various preliminary biochemical studies by various literatures. This plant is having the active phytoconstituents which are identified by the G Anburaj et al. The preliminary phytochemicals like carbohydrates, glycosides, alkaloids, steroids, Protein& Amino Acid, Tannins, Flavonoids, Saponins, Terpenoids, Fixed oil and fats and Gums and mucilage are the identified by using various phytochemical tests, this plant is showing the positive results of the various parts and the different extract having the different phytoconstituents. (Brahmam et al., 2015, Anburaj et al., 2016) And the bio-active components are identified by the GC-MS study the results of this study is confirmed the following active constituents present in the Tecoma stans Propane, 1,1,3-Triethoxy-5-Hydroxymethylfurfural, 1'-Hydroxy-4,3'-Dimethylbicycle, 9-Oxabicyclo[3.3.1] Nolan-2-One, 1,10-Decanediol, 1,2,3,4,7,7a-Hexahydro-2,4,7-Trimethyl-6H, Tropane, 2-Acetyl-2,3-Methylene-, 5-Undecanol, 2-Methyl, 6-Dodecanol, Silacyclopentane, 1,1-Dimethyl, Cyclobutanecarboxylic Acid, Decyl ester, Propanamide, 3-(1-Piperazinyl)-, Tetradecanoic Acid, Tetradecanoic Acid, Ethyl Ester, 2(4h)-Benzofuranone, 5,6,7,7atetr, L-(+)-Ascorbic Acid 2,6-

Dihexadecanoate, Hexadecanoic Acid, Ethyl Ester, N-Nonadecanol-1, 9,12-Octadecadienoic Acid (Z,Z), Ethyl (9z,12z)-9,12-Octadecadien, Octadecanoic Acid, 9,12-Octadecadienoate, N-Propyl 9,12,15-Octadecatrienoic Acid, Ethyl Ester, Octadecanoic Acid, Ethyl Ester and Hexatriacontane. The GC-MS study is identified the compounds by the retention time and the molecular mass of the compounds.(Lakshmi et., 2013, Kamilia et al., 2016)

# Active constituents with medicinal value of the *Tecoma stans*

- Tetradecanoic acid- Antioxidant, Lubricant, hypercholesterolemia, Cancer-preventive, Cosmetic
- Hexadecanoic Acid, Ethyl Ester- Antioxidant, hypocholesterolemic, Antiandrogenic, hemolytic, Alpha-reductase inhibitor.(Govindappa *et al.*, 2011, Sridharan *et al.*, 2014)
- I-(+)-Ascorbic acid 2,6-dihexadecanoate- Vitamin C, Antioxidant, Immunomodulator
- N-Nonadecanol-1- Anti-inflammatory, Hypocholesterolemic, Cancer preventive, Hepato-protective, Nematicide, Insectifuge Antihistaminic, Anti-arthritic, Anti-coronary, Antieczemic Antiacne, 5-Alpha-reductase inhibitor Antiandrogenic,
- 9,12-Octadecadienoic Acid (Z, Z)-Hypocholesterolemic, 5-Alpha-reductase inhibitor, Antihistaminic, Insectifuge, Antieczemic, Antiacne
- 9,12,15-Octadecatrienoic Acid, (Z, Z, Z)-Hypocholesterolemic, Nematicide Anti-arthritic, Hepatoprotective, Antiandrogenic, Nematicide 5-Alpha-reductase inhibitor, Antihistaminic, Anticoronary, Insectifuge, Antieczemic, Anticancer
- Octadecanoic acid- Cosmetic, Flavor, Hypocholesterolemic, Lubricant, Perfumery, Propecia, Suppository

# PHARMACOLOGICAL SCREENING OF TECOMA STANS

The *Tecoma stans* is having the many medicinal properties, the researchers are studied its medicinal properties by the *in vitro* and *in vivo* pharmacological evaluations. The current updates on the pharmacological screening are shown in the table.1 and table.2.

*In vitro* studies are performed by using particular parts of an organism, for example cells, microorganisms or biological molecules. The *in vitro* studies of the *Tecoma stans* are the shown in the table.1.

#### **Anti-Proliferative Activity**

Cancer is an awful sickness which is more than 100 different types and is categorized by uneven proliferation of the cells which required multidimensional approach for its treatment, control, prevention and is a second leading cause of death worldwide. Breast cancer is one of the longlasting ailments which may familiarity by females (32.1%) throughout her lifetime and is most commonly identified cancer in them.(Indra et al., 2010) The in vitro antiproliferative activity of the various parts of the Tecoma stans is done in the Breast cancer- MCF-7 cell lines by MTT assay. The Stem, Root, Bark and flowers extracts showed significant anti-proliferative action on the cell lines (MCF-7) but extreme action was found to be in extract stem bark of Tecoma stans.

#### **Antioxidant Activity**

Free radicals are having one or more unpaired electrons produced in pathological cell metabolism. The commonly produced free radicals are hydroxyl, superoxide, peroxyl radicals. These are the radicals are produced by the oxidation process in the system. Antioxidants are the used to secure the human body from reactive oxygen species. Broadcast of numerous bioactive compounds from plants has leads to the detection of new medicinal drug which have effective protection and treatment roles in against different diseases.(Shanmukha et al., 2014) The antioxidant activity of the plant is evaluated by the FRAP assay and the APTS, DPPH radical assays. These are the assays are then compared with the standard antioxidant drugs like vitamin C, Vitamin E etc.

#### Anti-microbial

The microbial infections are the produces the many of the health problems in the world. The using of synthetic antimicrobial drugs and the antibiotics may produce the some of the adverse effects so that the using of plant source as the antimicrobial agents is important to healing of microbes. In the plant extracts are undergoes the antimicrobial assays by disc diffusion method and the agar medium and minimum inhibitory concentration, etc., in this method the positive and negative strains of organisms and fungal strains are used to the microbial growth and this is inhibited by the adding of the drugs the zone of inhibition was measured and compared with the standard and determination of the antimicrobial activity.

#### **Hemolytic activity**

Hemolysis is the process of damage cytoplasmic membrane and producing cell lysis and death. The hemolytic activity is determined by the following procedure. Human blood of different groups (A, B, O) is collected from healthy volunteers in tubes containing heparin anticoagulant. And centrifuged at 3,000 rpm for 3 minutes and the hRBCs were collected. The cells are washed with PBS solution repeated until the supernatant was colorless. The hemolytic assay was performed in a microwell plate. The PBS is filled in each well. And wells added with ABCs. The serially diluted peptide solutions are added in the suitable wells. The hRBCs is used as negative control and hRBCs in Triton is used positive control. Then incubated for 1hour, the button formation of the wells is observed.(Amad et al., 2012)

#### Anti-Lipoxygenase activity

Anti-Lipoxygenase activities are studied by using enzyme-substrate complex concept. In this study linoleic acid (substrate) and lipoxidase (enzyme). Test drugs are dissolved in borate buffer pH 9.0 and added lipoxidase enzyme solution and incubated for 5 min at 250C. After incubation added with the linoleic acid solution, mixed well and absorbance was measured. (Kameshwaran *et al.*, 2014) Standard drug indomethacin was used. A dose-response curve was designed to decide the IC50 values.

#### Xanthine oxidase assay

Xanthine oxidase is the one of enzyme generates the reactive oxygen species. These are the xanthine oxidase are inhibited by some of the drugs and the determination of the inhibitory action of xanthine oxidase by xanthine oxidase assay. This assay was performed by the following procedure. Xanthine oxidase assay was evaluated spectrophotometrically. In this study the mixture of xanthine, xanthine oxidase and extract with phosphate buffer are incubated in a cuvette. The activity enzyme was articulated as the increase in absorption at 300 nm per unit time.

# Acetylcholinesterase Inhibitory activity

A cholinesterase or choline esterase is the enzymes which hydrolysis the acetylcholine to choline and acetate. These are the enzymes are inhibited by the choline esterase inhibitors, the acetylcholine enzyme inhibitors activity was performed by the following method. The AChE inhibitory assay was performed by the following procedure. The mixing of Tris-HCl, BSA buffer, extracts liquefied in buffer-methanol and Acetyl-cholinesterase.(Govindappa *et al.*, 2011) The above mixture was then incubated at room temperature for 2 min before the added with DTNB (5,5 Vdithiobis [2-nitrobenzoic acid], substrate acetylthiocholine iodide (ATCl). The development of yellow color was measured at 405 nm after 4 min.

s.	Activity	Part of the	Type of	Method	Main	Evaluation	Reference
no		plant	extract		materials	parameters	
1	Anti-	root, stem	ethanol	MTT Assay	Breast cancer-	Cell viability	Thirumal <i>et</i>
	Proliferative	bark and	(60°C) by		MCF-7 cell	(%),	al., 2013
	Activity	flowers	using a		lines,	Cell death	
			Soxhlet		Fetal bovine	(%)	
			apparatus		serum (FBS),		
					penicillin (100		
					U/ml), and		
					streptomycin		
2	Antioxidant	leaves	methanol,	DPPH radical	2,2-diphenyl-	%	Minal Wani
	Activity		ethanol,	scavenging	2-	scavenging	et al., 2014
			ethyl	activity	picrylhydrazyl	activity	
			acetate,		hydrate		
			and water		(DPPH),		
					ascorbic acid		
3	Antimicrobial	leaves	Hydro-	Disc	I wo-gram +ve	the	Kamilia et
	and antioxidant		distillation	diffusion	bacteria and	diameter of	al., 2016
			in a	method,	two gram –ve	zones of	
			Clevenger	Ine	bacteria,	innibition,	
			apparatus	Iviinimum Isekikitese	Fungi,	<i>%</i>	
				Concentratio	Amphotericin	scavenging	
					D. D. Vitamin	activity	
				DPPH radical	E		
				scavenging	L		
				activity			
4	Antimicrobial.		ethanol.	Paper disc	Two Gram	Diameter of	Govindappa
	Antioxidant		methanol.	method.	positive	zone of	et al 2011
			and water	FRAP assav.	bacteria. six	inhibition.	
				DDPH radical	Gram-	%	
				assay	negative	scavenging	
				,	bacteria,	activity	
					Fungi,		
					Chlorampheni		

Table 1: *in vitro* screening of *Tecoma stans* with current updates

					col, carbendazim, FRAP reagents, DPPH, L-ascorbic acid and BHT		
5	Antifungal and hemolytic activities	powered	chloroform and 70% ethanol	Drop diffusion method, Minimum Inhibitory Concentratio n method, Hemolytic Test	Fungi, Ketoconazole,	zone of inhibition	Ramesh <i>et</i> <i>al.,</i> 2009
6	Anti- Inflammatory, Lipoxygenase, Xanthine Oxidase And Acetylcholineste rase Inhibitory	powder	ethanol, methanol, and water	Alpha- reductase of albumin denaturatio n, Membrane stabilization test, Xanthine oxidase assay	Trypsin, the lipoxidase enzyme, xanthine oxidase, Aspirin, Acetyl- thiocholine iodide	Percentage inhibitory activities,	Govindappa <i>et al.,</i> 2011
7	Insecticidal	leaves	Methanol, water, Petroleum ether, Chloroform	Anti-Feedant Properties, Repellant Action	maize weevil, Beans weevil, Sugar	Percentage mortality rate	Tas <i>et al.,</i> 2015
8	Antimicrobial	heartwood	water, ethanol, methanol	Paper disc method	bacteria and fungi, chlorampheni col, carbendazim	Zone of inhibition (in mm)	Kottai <i>et al.,</i> 2012
9	Antibacterial	Roots	Methanol	Agar cup plate method	Bacteria, Ciprofloxacin	Zone of inhibition	Ramesh <i>et</i> al., 2009
10	Antibacterial	leaves	acetone, ethanol, chloroform , Diethyl ether and ethyl acetate	single disc diffusion method	Bacteria	Zone of inhibition	Subalakshm i <i>et al.,</i> 2017 Boonathi et
1 1 1	AITTILICTODIAL	ieaves	Ethanol,	agai	Dacterial and	Zone of	BOOPAth et

#### PRINT ISSN: 2394-6679 | E-ISSN: 2347-7881

			petroleum ether	diffusion method	fungal strain,	inhibition	al., 2017
12	Antioxidant And Antimicrobial	flowers	ethanol	DPPH radical scavenging assay, β –carotene bleaching assay, Reducing power assay, agar-well diffusion method.	β-carotene- linoleic acid, DPPH, potassium ferrocyanide, ferric chloride, bacterial and fungal strain	Scavenging effect (%), Zone of inhibition	Rajamuruga n <i>et al.,</i> 2013
13	antimicrobial	bark	Ethanol, water	Disc diffusion method	Bacterial and fungal strain Tetracycline, ampicillin	Zone of inhibition	Anburaj <i>et</i> <i>al.,</i> 2016
14	antibacterial activity	leaves	n-hexane		bacterial strains, Amikacin, Ampicillin	Zone of Inhibition (mm)	Sundas <i>et</i> <i>al.,</i> 2016
15	Antioxidant	heartwood	water, ethanol, and methanol	FRAP assay, DPPH activity	FRAP reagents, DPPH, L-ascorbic acid and BHT	Scavenging effect (%),	Kottai Muthu <i>et</i> <i>al.,</i> 2012
16	antimicrobial	leaves	water	agar disc diffusion method	Bacterial strains	Zone of inhibition (mm)	Senthilkum ar <i>et al.,</i> 2010

# IN VIVO PHARMACOLOGICAL SCREENING OF TECOMA STANS

*In vivo* are the effects of various drugs are tested on whole, living organisms or cells, generally animals, including humans, and plants. The *in vivo* studies of the *Tecoma stans* are shown in Table.2.

# Anti-inflammatory Activity

Inflammation is formed by the tissue reaction to infection, foreign substances or irritation. The inflammation process is important in the healing of wounds. Several mechanisms are involved in the inflammatory reactions such as the release of histamine, bradykinin, and prostaglandins. The antiinflammatory drugs are the used to prevent the inflammation. (Gharib *et al.*, 2007, Das *et al.*, 2010) These are the anti-inflammatory drugs are screened by the *in vivo* animal models and determination of the activity of the drug by the comparison with the standard and control groups.

#### **Central Analgesic Activity**

Pain is the symptom of numerous illnesses needful action with analgesics. The analgesic activity is the relives the pain symptom. These are the drugs are then screened by the *in vivo* animal models like HAFFNER's tail clip method in mice, Tail flick or other radiant heat methods, Tail immersion tests, Hotplate methods in mice or rats, Electrical stimulation, Monkey shock titration and Formalin test in rats.(Mohamed *et al.*, 2016)

#### Anticancer

The cancer cells are the having the properties of Uncontrolled growth, tissue invasion, and distinct metastasis. The Chemotherapy is used for the treatment of cancer, but they are greatly toxic, and

43

the negligible quantity of injected drug only can reach the cancerous tissue, may damage the normal system particularly bone marrow, epithelial tissue, reticuloendothelial system, and gonads. Now a day the plant medicine is the used to the treatment of cancer, the *Tecoma stans* are having the anti-cancer activity and which is evaluated by the WETS on survival time in the animal model.

#### **Cardio-protective effect**

Cardiovascular diseases (CVD) cause death in many countries. Myocardial infarction is the imbalance oxygen supply to the myocardium followed by the development of myocardial necrosis. The increased the toxic reactive oxygen species (ROS) such as O2-, H2O2, OH- etc. exerts simple oxidative pressure on myocardium prompting to CVD, for example, ischemic heart disease, atherosclerosis, congestive heart. The *Tecoma stans* are having the cardioprotective effect which is screened by the animal model and estimation of the antioxidant activities of the myocardium.

#### **Genotoxic study**

Genotoxicity is the property of chemical substances that damage the genetic information within a cell causing mutations, lead to cancer. All mutagens are genotoxic, but not all the genotoxic substances are mutagenic. The *Tecoma stans* plant is undergoing the genotoxic study and shown in table.2.

# Cytotoxicity study

Cytotoxicity is toxic to cells. The cytotoxicity of *Tecoma stans* in human hepatoblastoma was determined by incubating the cells up to 72-hours and changing with concentrations of herbal extracts. Toxic effects of *Tecoma stans* were originated to be attentiveness and time-dependent in the presence and absence of fetal bovine serum.

# Wound healing activity

Wound healing is the processes of growth and regeneration of wound tissue. The aim of wound repair is to promote wound healing in the shortest time possible, with least pain, discomfort, and scarring to the patient. Some medicinal plants are used in traditional medicine for wound healing. The *in vivo* study of the wound healing by the various wound animal models and compare with the standard group and control.

# Anti-hyperglycemic

Diabetes mellitus is an endocrine syndrome which mostly increases glucose level in the blood due to the defect in the insulin secretion or insulin action or both. In diabetes, the defect in the pancreas islet cells followed by insulin deficiency and causes diabetes. The anti-diabetic drugs are the used to the treatment of diabetes, these are the drugs are screened by the diabetic animal models such as chemical induced diabetes model, genetic induced, hormone induced and the viral-induced animal models. The parameters like glucose level were measured in this model. The *Tecoma stans* having the anti-diabetic activity which is evaluated by the chemical induced diabetes model.

#### **Protect CNS against Oxidative Damages**

The specific receptor binding of three neurotransmitters: GABA, an inhibitory transmitter and acetylcholine and glutamate. The oxidative effect produces the free radicals in the neurotransmitters and followed by causes of CNS. Anti-oxidative substances are protected central nervous system from oxidative effect. Recent studies are used herbal extracts as anti-oxidative agents; the Tecoma stans are having this property and protect the CNS.

# **Gastric Ulcer Healing Activity**

Gastric ulcers are the erosion of the gastric mucosa in the gastrointestinal tract. There are many factors can induce the ulcer like stress, alcohol, drugs, etc., numerous drug classes are used to the treatment of gastric ulcers, such as proton pump inhibitors, M1receptor blockers, and H2-receptor antagonists. The anti-ulcer drugs are screened by the various animal models. These are the animal models are the ulcer induced animal models there are many induction methods are available like stress, alcohol, and drugs (paracetamol). The measurement of the ulcer index and histopathological the studies bv the determination of the activity and compared with control and standard groups.

s.n	Activity	Part of	Animals model	Animal	Standard drug	Evaluation	Reference
ο		the plant		used		parameters	
		and					
		extract solvent					
1	Anti-	Ethanol/	Carrageenan-	Wistar	Indomethacin	the volume of	Brahmam
	inflammator	Flower	Induced Paw	rats of	does 20mg/kg	the paw (using	<i>et al.,</i> 2015
	У		Oedema	either	s.c.,	Digital	
				SEX (180-		plethysmometer	
2	Central	Ethanol	Hot plate	Swiss	Pentazocin, 10	) licking of paws	Brahmam
-	Analgesic	Aqueous	method	albino	mg/kg, i.p.)	shaking or	et al., 2015
	Activity	/ Flower		mice of	0, 0, 1,	jumping off the	,
	·			either		surface (Eddy's	
				sex (25-		hot plate)	
				30gms)			
3	Antinocicept	Alcohol	Hot plate	Albino	Pentazocine(10	licking of paws,	Lakshmi et
	ive activity	and	method,	mice and	mg/kg)., i.p,	shaking or	al., 2013
	and Anti-	water/	Formalin-	wister	Pentazocine	Jumping off the	
	v activity	Leaves	licking model	either	(1011g/kg) i.p. Diclofenac	bot plate)	
	yactivity		Acetic acid	sex	sodium (10	%inhibition of	
			induced	(weighin	mg/kg, p.o.	Paw licking,	
			writhing test,	g 18-24	0, 0, 1	% inhibition of	
			Carrageenan-	g, 150-		writhing	
			induced rat	200 g		response.	
			paw edema,			% inhibition of	
						paw edema	
4	A	\Aletaulaa		Curies		Inflammation	Cuidheanan
4	Anticancer	water/ae	WEIS ON	SWISS	5- Fluorouracii	Tumor volume,	Sridnaran
		nai parts	Survival time	mice	(2011g/kg.bw.)	count and	et ui., 2014
				mille		Viable and non-	
						viable cell	
						counts	
5	Cardio-	Ethanol/	isoproterenol-	Wister	Simvastatin (60	ALT,	Shanmukh
	protective	Flowers	induced	rats (150-	mg/kg)p.o	AST, LDH, CK,	a et al.,
	effect		myocardial	250g)		TC, TG, LDL and	2014
			infarction			depletion of	
						ADL levels	
						neroxidation	
						SOD, and CAT,	
6	Genotoxic	Water/Le	Bone marrow	Male		Mitotic index,	Amad M et
	and	aves	chromosome	BALB/c		Chromosome	al., 2012
	cytotoxic		assay,	mice (22-		abbreviation,	
			Mitotic index	24g)		Percentage	
			assay,			INHIBITION OF	
			chromosome			IVIEF	

Table 2: In vivo screening of Tecoma stans with current updates

	I			1			
7	Wound healing	Ethanol/ Flowers	abbreviation assay, Mouse embryo fibroblast, Viable cell count Excision wound model, Incision wound model, Burn wound model	Wister albino rats (150- 180g)	Silver sulfadiazine (0.01%)	Percentage wound concentration, Epithelialization period, Tensile strength, Hydroxyproline, Histo- pathological study	Kameshwa ran <i>et al.,</i> 2014
8	Wound healing	Petroleu m ether, Chlorofor m, Methano I/ Bark	Excision wound, Incision wound	Wister albino rats (180- 250g)	Vaseline, 2% gum acacia suspension (1 ml/kg,p.o).	Percentage wound concentration, Epithelialization period	Das <i>et al.,</i> 2010
9	Anti-cancer	Methano I/ Flowers	Antitumor activity, Hematological parameters.	Male Swiss albino rats (150- 180g)	5- Fluorouracil (20mg/kg.bw.)	Tumor volume, Tumor weight, Viable/non- viable cell count, mean survival time, Body weight, food intake	Kameshwa ran <i>et al.,</i> 2012
10	Antispasmo dic	Alcohol/ Leaves	Bio-assay	Male adult Wister rats (200- 250g)	carbachol, tetraethylamm onium, propranolol, naloxone, glibenclamide	Percentage relaxation, Percentage contraction	Gharib <i>et</i> <i>al.,</i> 2007
11	Anti- hyperglyce mic	Methano I/ Leaves	STZ induced diabetes model	albino rats (100-150 g)	metformin hydrochloride( 500 mg/kg)	Carbohydrate tolerance curve, TC, TG, HDL, glucose, creatinine, uric acid, ALT levels,	Taher <i>et</i> <i>al.,</i> 2016
12	Protect CNS	Ethanol/ Flowers	Tail-suspension Test, Forced- swimming Test, Actophotomet er	Wistar albino rats (200- 300g)	No standard group only control group treated with vehicle	immobility time, locomotors activity	Kameshwa ran S <i>et al.,</i> 2014

PharmaTutor

# PRINT ISSN: 2394-6679 | E-ISSN: 2347-7881

13	Gastric Ulcer Healing	Petroleu m ether/ Leaf	Pylorus Ligation Induced Ulcers	Wister albino rats (150- 180g)	Ranitidine	Volume of Gastric juice ml, pH, Free acid (meq/l), Ulcer score %	Arnabadity a <i>et al.,</i> 2012
14	Hepatoprot ective	Petroleu m ether, Chlorofor m, Ethanol/L eaves	Thioacetamide induced hepatotoxicity, CCL4 induced nephrotoxicity	Wistar albino rats (150- 250g)	Silymarin 100 mg/kg p.o.,.	liver weight, liver volume, biochemical markers, GSH, lipid peroxidation levels,	Shanmukh a <i>et al.,</i> 2013
15	Nephroprot ective	Petroleu m Ether, Chlorofor m, Ethanol, Water/ Leaves	Cisplatin- induced nephrotoxicity in rats, Gentamicin- induced nephrotoxicity in rats, Paracetamol- induced nephrotoxicity in rats	Wister albino rats (weighin g 150- 250g) and albino mice (weighin g 20-25g)	saline 1ml/kg p.o	blood urea, serum creatinine, kidney weight, GSH levels, body weight, lipid peroxidation levels	Shanmukh a <i>et al.,</i> 2012

# CONCLUSION

*Tecoma stans* are having the many of the active phytoconstituents which are leads to the great medicinal value of this plant. In this review was concluded that the various parts of the plants are having various pharmacological actions like anti-inflammatory, analgesic, anticancer cardio-protective effect, genotoxic, cytotoxicity, wound healing, anti-hyperglycemic, protect CNS, gastric ulcer healing, antiproliferative, antioxidant, anti-microbial, hemolytic activity, anti-lipoxygenase and acetyl-cholinesterase inhibitory activities. And this review was used to develop the further research in this plant *Tecoma stans*.

**ACKNOWLEDGEMENT:** I thankful to the almighty for blessings in a successful completion of this work, my special thanks to KMCH College of pharmacy, and I would like to thank my friends V.Sanish Devan, M.Sanjay, A.Jeevanantham, M.Thanga Kokila, J.Kanu Priya and V.Sri Vidya who has helped me to gather fine details on this work.

# **↓** REFERENCES

1. Amad M al-Lazzari (2012); Genotoxic and Cytotoxic study of Tecoma stans Bignoniaceae; Pakistan journal of biological sciences; Vol. 15 No. 2; 92-97

2. Anburaj G., Marimuthu M. and Manikandan R (2016); In vitro antimicrobial activity of aqueous and Ethanol extracts of Tecoma stans bark against pathogenic Bactria; International Recent Research Journal on Science and Technology; Vol. 8 No. 2; 26-28

3. Anburaj G., Marimuthu M., Rajasudha V. and Manikandan R (2016); Phytochemical screening and GC-MS analysis of ethanolic extract of Tecoma stans (Family: Bignoniaceae) Yellow Bell Flowers; Journal of Pharmacognosy and Phytochemistry; Vol. 5 No. 4; 172-175

4. Anburaj G., Marimuthu M., Sobiyana P. and Manikandan R. (2016); A Review on Tecoma stans; International

Vol. 6, Issue 1 | magazine.pharmatutor.org

47

Journal of Engineering Research and Modern Education; Vol.1 No.1; 43-49

5. Arnabaditya Mohanty, Vinod Kumar Sahu, Ashutosh Mishra, Dusmanta Kumar Pradhan and Manas Ranjan Mishra (2012); Gastric ulcer healing activity of Tecoma stans Leaf; International Research Journal of Pharmaceutical Sciences; Vol. 3 No.1; 32-33

6. Boopathi T., Gopalasatheeskumar K., Parthiban S., Sangeetha G., Thanga Kokila M. and Manimaran T (2017); Evaluation of Antimicrobial Activity of Tecoma stans and Muntingia calabura; World Journal of Pharmaceutical Research; Vol. 6 No. 3; 617-627

7. Brahmam B., Sirisha K., Sathish Kumar M., Narendra Babu A. and Rama Rao N.V (2015); Evaluation of Central Analgesic Activity of Tecoma stans Flower Extracts; Vol. 4 No. 1; 89-92

8. Brahmam B., Sirisha K., Sathish Kumar M., Narendra Babu A., Rama Rao N.V. and Rama Rao N (2015); Evaluation of Anti-inflammatory Activity of Flower Extracts of Tecoma stans on Carrageenan-Induced Paw oedema in Rats by Using Digital Plethysmometer; Research Journal of Pharmaceutical, Biological and Chemical Sciences; Vol. 6 No. 5; 641- 644

9. Chaugan S.V.S., Jolly Singh and Satoshi Tahara (2004); Role of phenolic sans boron in reproductive success in seasonally transient sterile Tecoma stans L; Indian journal of experimental biology; Vol. 42 No. 1; 197-201

10. Das C., Dash S., Sahoo D.C. and Mohanty A (2010); Evaluation of Methanolic Bark Extract of Tecoma stans Linn, for Wound Healing in Albino Rats; International Journal of Pharmacy and Technology; Vol. 2 No. 3; 735-742 11. Divya Sri G., Narendra Babu A., Sathish Kumar M., Venkateswarlu V. and Ashok Kumar K (2014); Pharmacognostical Characteristics and Medicinal Uses of Tecoma stans: A Review; Journal of Medical and Pharmaceutical Innovation; Vol. 1 No.2; 1-4

12. Gharib Naseri M.K., Asadi Moghaddam M. and Bahadoram S (2007); Antispasmodic effect of Tecoma stans (L.) Juss leaf extract on rat ileum; DARU; Vol. 15 No. 3; 123-128

13. Govindappa M., Sadananda T.S., Channabasava R. and Vinay B Raghavendra (2011); In vitro Anti-Inflammatory, Lipoxygenase, Xanthine Oxidase and Acetylcholinesterase Inhibitory Activity of Tecoma stans (L.) Juss. Ex Kunth; International Journal of Pharma and Bio Sciences; Vol. 2 No. 2; 275-285

14. Govindappa M., Sadananda T.S., Channabasava R., Jeevitha M.K., Pooja K.S., Vinay B. and Raghavendra (2011) Antimicrobial, Antioxidant Activity and Phytochemical Screening of Tecoma stans (L.) Juss. Ex Kunth; Journal of Phytology Phyto-pharmacology; Vol. 3 No. 3; 68-76

15. Indra Gandhi M. and Ramesh S (2010); Antifungal and hemolytic activities of organic extracts of Tecoma stans (Bignoniaceae); Journal of Ecobiotechnology; Vol. 2 No.2; 26-32

16. Kameshwaran S., Senthilkumar R., Thenmozhi S. and Dhanalakshmi M (2014); Wound healing potential of ethanolic extract of Tecoma stans flowers in rat; Pharmacologia; Vol. 1 No. 1; 215-221

17. Kameshwaran S., Sundaraganapathy R., Thenmozhi S., Dhanalakshmi M., Vasuki K. and Manjuladevi K (2014); Tecoma stans protect Central Nervous System Against Oxidative Damages of Electromagnetic Radiation on Rat; Acta Biomedica Scientia; Vol. 1 No.1; 40-44

18. Kameshwaran S., Suresh V., Arunachalam G., Kanthlal S.K. and Mohanraj M (2012); In vitro and in vivo anticancer activity of methanolic extract of Tecoma stans flowers; International research journal of pharmacy; Vol. 3 No. 3; 246- 251

19. Kamilia F. Taha, El-sayeda A. El-kashoury, Shahira M. Ezzat and Naglaa A. Saleh (2016); Antimicrobial and antioxidant activity of volatile constituents of the leaves of Tecoma Smithii Will Wats; Global Journal of Medicinal Plant Research; Vol. 4 No. 4; 16-22

20. Kottai Muthu A., Borse L.B., Thangatripathi A. and Borse S.L (2012); Antioxidant activity of heartwood of Tecoma stans. (L) Juss. Ex Kunth; Journal of Pharmacy Research; Vol. 5 No. 2; 896-898

21. Kottai Muthu A., Laxmikant B. Borse, Thangatripathi A. and Sandhya L. Borse (2012); Antimicrobial Activity of Heartwood of Tecoma stans; International Journal of Pharmacy and Pharmaceutical Sciences; Vol. 4 No. 3; 384-386

22. Lakshmi Prasanna V., Lakshman K., Medha M. Hegde and Vinutha Bhat (2013); Antinociceptive and Anti-Inflammatory activity of Tecoma stans Leaf Extracts; Indian Journal of Research in Pharmacy and Biotechnology; Vol. 1 No. 2; 156-160 23. Mohamed Abdel-Hamid Taher, Dawood Hosni Dawood, Mostafa Ibrahim Sanad and Ramadan Ahmed Hassan (2016); Searching for anti-hyperglycemic phytomolecules of Tecoma stans; European Journal of Chemistry; Vol. 7 No. 4; 397-404

24. Name H. and Minal Wani (2014); Callus Induction Studies and Active Components and Antioxidant Activity Investigation from Leaves and Callus of Tecoma stans L. Juss. Ex Kunth; Research Journal of Pharmaceutical, Biological and Chemical Sciences; Vol. 5 No. 2; 604-610

25. Rajamurugan R., Thirunavukkarasu C., Sakthivel V., Sivashanmugam M. and Raghavan C.M (2013); Phytochemical Screening, Antioxidant and Antimicrobial Activities of Ethanolic Extract of Tecoma stans Flowers; Int J Pharm Bio Sci.; Vol. 4 No. 2; 124-130

26. Raju S., Kavimani S., Uma Maheshwara Rao V. and Sreeramulu Reddy K (2011); Tecoma stans (L.) Juss. ex Kunth (Bignoniaceae): Ethnobotany, Phytochemistry and Pharmacology; Journal of Pharmaceutical and Biomedical Sciences; Vol. 8 No. 7; 1-5

27. Ramesh T., Anusha V. and Ravi Kumar (2009); Antibacterial Activity of Methanolic Extract of Roots of Tecoma stans; Int. J. Chem. Sci.; Vol. 7 No. 1; 6-8

28. Senthilkumar C.S., Suresh Kumar M. and Rajasekara Pandian M (2010); In vitro Antibacterial Activity of Crude Leaf Extracts from Tecoma stans (L) Juss. Et Kunth, Coleus Forskohlii and Pogostemon Patchouli against Human Pathogenic Bacteria; International Journal of PharmTech Research; Vol. 2 No. 1; 438-442

29. Shanmukha I., Abubaker Siddiq, Prabhu K. and Ramachandra Setty S (2012); Effect of Tecoma stans Leaves Extract on Experimentally Induced Renal Injury In Various Animal Models; Am. J. PharmTech Res.; Vol. 2 No.6; 800-809

30. Shanmukha I., Vijay Kumar M. and Ramachandra Setty S (2013); Effect of Tecoma stans Leaves for its Preventive Role on Experimentally Induced Liver Toxicity; International Journal of Pharm Tech Research; Vol. 5 No. 3; 915-923

31. Shanmukha I., Vijay Kumar M. and Ramachandra Setty S (2014); Cardioprotective effect of hydroalcoholic extract of Tecoma stans flowers against isoproterenol-induced myocardial infarction in rats; Asian Pac J Trop Dis.; Vol. 4 No. 1; 378-384.

32. Sridharan G., Sarvanan R. and Brindha P (2014); Evaluation of Anticancer Potentials of Tecoma stans (L). Juss.Ex. Kunth against EAC Cell Lines; International Journal of Pharmacy and Pharmaceutical Sciences; Vol. 6 No. 1; 88-92

33. Subalakshmi T. and Jepa Chandra Mohan (2017); Inhibitory Effect of Different Solvent Extracts of Tecoma stans, Ixora Coccinea and Aerva Lenata Leaves on Pseudomonas aeruginosa and Streptococcus Sp. of cattle Pathogens; World Journal of Pharmacy and Pharmaceutical Sciences; Vol. 6 No. 2; 1219-1228

34. Sundas Iltaf, Zaheer-Ud-Din Khan, Rizwana Rafique and Anjum Parveen (2016); Evaluation of antibacterial activity of leaf extracts of Mansoa alliacea (Lam.), Tecomaria capensis (Thunb.) Spach and Tecoma stans (L.) Juss. Ex; Journal of Biodiversity and Environmental Sciences; Vol. 9 No. 1; 69-75

35. Sunitha Katta, Ganapathy Seru and Sridhar Y (2016); Constituents from the Leaves of Tecoma stans Juss; World Journal of Pharmaceutical Sciences; Vol. 4 No. 12; 272-274

36. Sunita Verma (2016); Phytochemical and pharmacological review study on Tecoma stans Linn; Journal of Medicinal Plants Studies; Vol. 4 No. 5; 162-164.

37. Tavs A. Abere and Comfort O. Enoghama (2015); Pharmacognostic standardization and insecticidal activity of the leaves of Tecoma stans Juss (Bignoniaceae); Journal of Science and Practice of Pharmacy; Vol. 2 No. 1; 39-45 38. Thirumal M., Kishore G. and Surya Srimanthula (2013); Anti-Proliferative Activity of Various Parts of Tecoma stans (L.) Against Human Breast Cancer Cells In vitro; Research Journal of Pharmaceutical, Biological, and Chemical Sciences; Vol. 4 No. 2; 305-313